International Journal of Agricultural Science and Research (IJASR) ISSN(P): 2250-0057; ISSN(E): 2321-0087

Vol. 5, Issue 2, Apr 2015, 33-40

TJPRC Pvt. Ltd.



A REVIEW: ROLE OF PHOSPHOROUS SOLUBILISING AND SULFUR

OXIDIZING BACTERIA IN MINE RECLAMATION

SHIKHA DIXIT¹ & APPU KUTTAN K. K.²

¹Department of Biological Science and Engineering, MANIT, Bhopal, Madhya Pradesh, India ²MANIT, Bhopal, Madhya Pradesh, India

ABSTRACT

Mining is a necessary evil leads to various types of environmental damages and deterioration of effected soil area. This makes the need of revegetation in the Affected area i.e. reclamation of mining area to make it suitable for further use. There are various methods to improve the physical, chemical and biological properties of soil from which use of biofertilizer is one of the cost effective and pollution free methods. The current review mainly focused on the role of phosphorous solubilizing and sulfur oxidizing bacteria as biofertilizer to regain the soil fertility in mining area.

KEYWORDS: Phosphorous Solubilizing Bacteria, Sulfur Oxidizing Bacteria, Biofertilizer, Bacteria

INTRODUCTION

Mines and Mining are the areas where ores for mining can be found. These may be above the ground or underground and scattered in fair number around gielinor. But mines are considered as necessary evil because mining leads to deterioration of soil quality and water pollution in affected areas. Major sources of erosion, sediment loading of mining sites can include open pit areas, heap and dump leaches, waste rock and overburden piles. A further concern is that, exposed materials from mining operations may contribute sediments with chemical pollutants, primarily heavy metals. Mining operations routinely modify the surrounding landscape by exposing previously undisturbed earth materials (Guide book for evaluating mining project EIA). So, there is need to reduce pollution and relcamation of mining area to make the mining area suitable for further use after mining.

Use of Biofertilizers in Reclamation of Mining Area

Since Mining results in loss of soil fertility, depletion of useful microbial communities, soil erosion and water quality deterioration, therefore there is an urgent need to develop technologies to reclaim these areas for further potential use. There are many methods to improve the physical, chemical and biological properties of soil from which revegetation is most popular and successful method(Sheoran et al., 2010). But this needs a good soil quality which can be improved by application of biofertilizers. Some of the beneficial microorganisms such as *Rhizobium*, *Azotobacter*, Phosphate solubilizing bacteria and blue green algae etc. can be used as biofertilizer in reclamation program. Soil microbes not only fix the nitrogen symbiotically but some microbes like phosphate solubilizing bacteria renders unavailable form of phosphorous to available form of phosphorous to reduce dependence on phosphorous from external sources. The soil microorganism also exhibit great metabolic versatility to make them suitable for growth in nutrient deficient soil. There are several evidence that suggest the prominent revegetation of disturbed sites when the seedlings of planted species are pre-inoculated with microbial inoculants (Adholeya et al., 1997; Bhatia et al., 1998; Aron, 2002; Fay et al., 1999; Franco et

al., 1997; Haselwandter and bowen, 1996; Mishra et al., 1991; Reddell et al.,1999; Setiadiet al.,2002; Sharma et al., 2001; Sharma et al., 1997). Use of biofertilizers may helpful in the development of a low cost technology to rehabilitate the mining areas and increase their productivity.

WHAT IS THE BIOFERTILIZER?

Biofertilizers are the products supplied with living cells of different types of microorganism having capacity to convert nutritionally important elements from unavailable to available forms through some specific biological processes. These biofertilizers can be applied to seeds or plant surface or soil(Rokhzadiet al.,2008). Beneficial microorganisms in biofertilizers have the ability to accelerate and improve plant growth and protect plants from pests and diseases (El-yazeid et al., 2007). The applicability of microbial inoculums is well known from the history which passes from generation to generations of farmers. It was recognized by the process of decomposition of organic residues and agricultural by-products by the culture organisms through different mechanisms and gives healthy harvest of crops (halim et al., 2009). The microbial inoculants were started in the late 1940's at Malaysia and peaking up in 1970's with the use of *Brady rhizobium* inoculation on legumes. Government research institute, the Malaysian Rubber Board (MRB) had been conducting research on *Rhizobium* inoculums for leguminous cover crops in the inter rows of young rubber trees in the large plantations.

In general there are six major steps in biofertilizer making. These includes choosing active organism, isolation and selection of target organism, selection of carrier material, selection of propagation method, prototype testing and large scale testing. Biofertilizers are generally prepared as powder form of carrier based inoculants consists of effective microorganisms. This makes biofertilizerhandy, effective, stable and suitable for long term storage. Carrier must be properly sterilized by autoclaving or gamma irradiation. Various types of carrier material are available for seed or soil inoculation. If biofertilizer is to be produced in powder form then tapioca flour or peat the right carrier materials. Organism that are commonly used as biofertilizers comonstituents are nitrogen fixers, potassium solubilizer and phosphorous solubilizer, or with combination of the molds or fungi. Now a day some sulfur and iron oxidizing bacteria are also included as components of biofertilizers. This review mainly focused on phsfatesolubilising and sulfur oxidizing bacteria as biofertilizer and their role on reclamation of mining area.

Phosphate Solubilizing Bacteria

Phosphorus is the key element in many metabolic processes in plants such as photosynthesis, respiration, energy transfer, signaling etc (Khan et.al., 2010) and nitrogen fixation in legumes (Saber et al., 2005). Phosphorus is present in soil as inorganic and organic forms, but in the unavailable form of precipitated mineral complexes. These precipitated forms can not be absorbed by the plants (Rengel and Marschner, 2005). Only 0.1 % of the total phosphorus is present in a soluble form i.e. available to the plants (zhou et Al., 1992). The term phosphate fixation explains the reactions that remove available phosphate from the solution into the solid phase of soil (Barber et al., 1995). Phosphate fixation involves two reactions: (1) phosphate sorption on soil surface and (2) phosphate precipitation by free Al³⁺ and Fe³⁺. So there is need to solubilize this phosphate to make it available for the plants. The phosphate solubilizing bacteria are the promising organism for the same. The natural occurrence of phosphate solubilizing bacteria in soil was first evidence in 1903 (khan et al., 2007). There are some phosphate solubilizing fungus also present in soil but with less percentage i.e. 0.1 to 0.5 % in comparison to phosphate solubilizing bacterial presence of 1 to 50 % (chenn et al., 2006). The main ectorhizospheric bacteria are *Pseudomonas* and *Bacilli*, and endosymbiotic rhizobia have been considered as effective phosphate

solubilizers (Igual et al., 2001). Bacterial genera of *Pseudomonas, Bacillus, Rhizobium* and *Enterobacter* are the most powerfull PSB (Whitelaw et al., 2000). *Bacillus megaterium, Bacilluscirculans, Bacillus subtilis, Bacillus polymyxa, Bacillus sircalmous, Pseudomonas striata Enterobacter* be find as the most important bacterial strains (Subbarao et al., 1998; kucey et al., 1989). The mechanism of phosphate solubilization involves many different reactions mediated by PSB. PSB release metabolites such as organic acids, which chelate the cation bound to phosphate through hydroxyl and carboxyl groupsor lowers the pH or compete for the sorption to the soil surface bound (Nahas et al., 1996). This makes the phosphate free from cation and available to the plants (Hilda and fraga, 2000; Khiari and Parent, 2005). Some PSB can enhance plant growth by enhancing the availability of some trace elements such as iron, zinc etc. (Ngoc et al., 2006), synthesize some enzyme responsible for hormonal modulation in plants, may limit the available iron via siderophore production (Akhtar and Siddiqui, 2009). H⁺excretion originating from NH₄⁺ assimilation as proposed by Parks et al. could be an alternative mechanism of Phosphate solubilization. In contrast to expectations, an HPLC analysis of the culture solution of *Pseudompnass*p. Did not showed any organic acid even though solubilization happened (Illmerand and Schinner, 1995). They also stated that most probable reason of solubilization without acid production is the release of protons accompanying respiration or NH₄⁺ assimilation.

This potential of PSBs makes them the main component of Biofertilizers. The reclamation of mining area demands the increase in fertility of the soil of effected area by fulfilling mineral deficiency, which could be completed by using these PSB as biofertilizers. There is evidence of increase in growth performance of *Acacia nilotica*, *Acacia catechu*, *Buteamonosperma* and *Pongamiapinnata* by the application of PSB with the combination of blue green algae (Dubey K. et al., 2006). The PSB solubilize the fixed soil phosphorus and applied phosphate resulting in higher crop yields (Gull et al., 2004.)

Sulfur Oxidizing Bacteria

Sulfur is also one of the essential element required by plats for optimum growth. Sulfur occurs in a wide variety of organic and inorganic combinations i.e. unavailable to plants. The dominant form of sulfur taken up by plants is sulfate (VidyalakshmiR. et al., 2009). Sulfur transformations in soil are primarily done by microbial activities such as oxidation, reduction, immobilization and mineralization. The biological transformations of sulfur have been observed by Vernadskii et al. (1927). The sulfur oxidizing bacteria are mainly gram negative bacteria currently distributed as species of Thiobacillus, Thiomicrospira and Thiosphaera, but some heterotrophssuch as Paracoccus, Xanthobacter, Alcaligensand Pseudomonas can also show chemolithotrophic growth on inorganic sulfur medium (Kuenen et al., 1982). Based on metabolism, there are two groups of sulfur oxidizing bacteria: The obligate chemoliithotrophs, which can only grow when supplied with oxidisable sulfur compounds and heterotrophs that can also use the chemolithoautotrophic mode of growth. The both groups can use CO2 as carbon source. The obligate chemolithotrophs include Thiobacillusthioparus, dinitrificans(facultative neapolitanus, T. denitrifier), Thiobacillusthiooxidans(extreme acidophile), Thiobacillusferrooxidans(acidophilic ferrous iron-oxidiser), Thiobacillushalophilus(halophile) and some species of Thiomicrospira. heterotrophs include Thiobacillusnovellus, *T*. acidophilus (acidophile), The Thiobacillusaquaesulis(moderate thermophile), Thiobacillusintermedius, Paracoccusdenitrificans, versutus, Xanthobactertagetidis, Thiosphaerapantotrophand Thiomicrospirathyasirae.

Biological oxidation of hydrogen sulphide to sulphate one of the major reaction mechanism of sulfur cycle on earth. Other than hydrogen sulphide bacteria have the ability to oxidize sulfur, sulfite, thiosulfate and various polythionates

under alkaline (Sorokin et al., 2001) neutral or acidic conditions (Harrison, 1984). Aerobic sulfur oxidissing bacteria belongs to genera like *Acidianus* (Friedrich, 1998), *Acidithiobacillus* (Kelly et al., 2000), *Aquaspirillum* (Fredrich, 1981), *Aquifer* (Humber et al., 1999), *Bacillus* (Arango et al., 1991), *Beggiatoa* (Strohl et al., 1989), *Methylobacterium* (Kelly et al., 1990; De Zwart et al., 1996), *Paracoccus, Pseudomonas* (Friedrich et al., 1981), *Starkeya* (Kelly et al., 2000), *Sulfolobus, Thermithiobacillus* (Kelly et al., 2000), *Thiobacillus and Xanthobacter* (Freidrich et al., 1981) and are mainly mesophilic. Phototrophic anaerobic sulfur oxidizing bacteria mainly belongs to genera like *Allochromatium* (Imhoff et al., 1998), *Chlorobium, Rhodobacter, Rhodopseudomonas, Rhodovulum* and *Thiocapsa* (Brune, 1989). Some bacteria such as *Thiocapsaroseopersicina, Allochromatiumvinosum and Rhodopseudomonasacidopphila* (purple non sulfur bacteria) showed growth in dark (Siefert et al. 1979; Kondratieva, 1989).

Though sulfur may be oxidized by many groups of bacteria, but Waksman (1932) pointed out *Thiobacilli* as the most promising and characteristic group of bacteria performing the oxidative part of sulfur transformation in soil. Inoculation of *Thiobacilli* generally increases the rate of sulfur oxidation (Kapoor et al.,1989). This makes sulfur oxidizing bacteria to be used as biofertilizers. Rock biofertilizers from P and K rocks with sulfur inoculated with *Acidithiobacillus* have been applied to different regions of the rain forest zone and in the semiarid region of the Brazilian Northeast, and results shown were excellent.

Combination of Phosphate Solubilizing and Sulfur Oxidizing Bacteria as Biofertilizer Component

Researchers have found the value of sulfur oxidizing bacteria in enhancing the phosphorous availability to the plants when supplied in combination of rock phosphate with sulfur, organic matter, PSB and sulfur oxidizing bacteria (Chein et al., 1996; Vessey et al., 2003). The acidity of the soil, amount of soluble calcium and type of chelating legands are the basic parametersaffecting the phosphorous availability (Chein et al., 1996). The enhanced phosphorous availability in rock phosphate combined with elemental sulfur and *Thiobacillus*(Biospur) has been also observed by Stamford 2002. The growth and oil production of *Raphanussativus*was significantly increased when rock phosphate with *Thiobacillus*was used as biofertilizer(khatbirasool, 2011). Parameters affects the phosphorus availability of rock phosphate when used in combination with elemental sulfur include the type of rock phosphate, the ratio of rock phosphate to elemental sulfur and condition of soil and crop (Rajan S., 2002).

CONCLUSIONS

Bio fertilizers can play key role in reclamation of mining area to increase the soil fertility for the revegetation of the mining area. The phosphate solubilizers mixed with organic material, elemental sulfur and sulfur oxidizing bacteria can be proved as a good bio fertilizer for improving soil quality. Furthermore, biofertilizer can be a good replacement of chemical fertilizers in the terms of cost and reduction of environmental pollution.

REFERENCES

- 1. Abdul, Halim N. B. (2009). Effects of using enhanced biofertilizer containing N-fixer bacteria on patchouli growth. Faculty of Chemical and Natural Resources Engineering University Malaysia Pahang.p.145.
- 2. Adholeya, A., Sharma, M.P., Bhatia, N. P., Tyagi, C. (1997). Mycorrhizal Biofertilizers: a tool for reclamation and biofertilizer. Proceeding: National Symposium on Microbial Technology in Environmental Management and Resource Recovery, 1-2 October 1997. New Delhi.

- 3. Akhtar, M.S., and Siddiqui, Z.A. (2009). Effect of phosphate solubilizing microorganisms and Rizobium sp. on the growth, nodulation, yield and root-rot disease complex of chickpea under field condition. Afr. J. Biotech., 8(15), 3489-3496.
- 4. Anon, (2002). CBR Centre for Biotechnology Research, Effect of bio- organic on soil and plant productivity improvement of post tin mine site at PT Koba Tin Project Area, Bangka. Centre for Biotechnology Research, Bogor Agricultural University.
- 5. Aragno, M. (1991). Aerobic chemolithoautotrophic bacteria, In: Thermophilic bacteria. J.K. Kristjansson (Ed.) CRC Press, Boca Raton, Fla, 7-103.
- 6. Barber, S.A. (1995). Soil nutrient bioavailability. A mechanistic approach, Wiley, Newyork.
- 7. Bhatia, N.P., Adholeya, A., Sharma, A. (1998). Biomass production and changes in soil productivity during long term cultivation of *Prosopisjuliflora* inoculated with VAM and *Rhizobium* spp. In a semi-arid wasteland. Biology and Fertility of Soils, 26,208-214.
- 8. Brune, D.C. (1989). Sulfur oxidation by phototrophic bacteria. Biochem.Biophys.Acta, 975, 189-221.
- 9. Chein, S.H., Menon, R.G., Billingham, K. (1996). Phosphorus availability from phosphate rock as enhanced by water soluble phosphorus. Soil Sci. Soc. Am. J., 60, 1173-1177.
- 10. Chen, Y.P., Rekha, P.D., Arun, A.B., Shen, F.T., and Lai, W.A. et al. (2006). Phosphate solubilizing bacteria from subtropical soil and their tricalcium phosphate solubilizing activities. Appl. Soil Sci., 34, 33-41.
- 11. De Zwart, J.M.M., Nelisse, P.N., and Kuenen J.G. (1996). Isolation and characterization of *Methylophagasulfidovorans*, sp. nov.anobligatelymethylotrophic,aerobic, dimethyl sulfide oxidizing bacterium from amicrobial mat. FEMS Microbiol. Ecol., 20, 261-270.
- 12. Dibey, K., Singh, V.K., and Mishra, C.M. et al. (2006). Use of Biofertilizer for Reclamation of Silica Mining Area. Billings land reclamation symposium. 176-181.
- 13. El-Yazeid, A.A., Abou-Aly, H.A., Mady, M.A., and Moussa, S.A.M. (2007). Enhancing growth, productivity and quality of squash plants using phosphate dissolving microorganisms (bio phosphor) combined with boron foliar spray. Res. J. Agric. Biol. Sci., 3(4), 274-286.
- 14. Fay, D.A., Mitchell, D.T., Parkes, M.A. (1999). A preliminary study of the mycorrhizal association of tree seedlings growing on mine spoil at Avoca, Co. Wicklow. Biology and Environment: Proceedings of the Royal Irish Acad., Section B, 99,19-26.
- 15. Franco, A.A., Faria, S.M.-de, Faria, S.M. (1997). The contribution of N₂ Fixing tree legumes to land reclamation and sustainability in the tropics. In international symposium on Sustainable agriculture for the tropics: the role of biological nitrogen fixation, Angra dos Ries, Rio de Janerio, Brazil, 29, 5-6.
- 16. Friedrich, C.G. (1998). Physiology and genetics of sulfur-oxidizing bacteria. Adv. Microb. Physiol., 39, 235-289.
- 17. Friedrich, C.G. and Mitrenga G. (1981).Oxidation of thiosulfate by *Paracoccus denitrificans* and other hydrogen bacteria.FEMS Microbiol.Lett., 10, 209-212.

- 18. Gull, M., Hafeez, F.E., Saleem, M., and Malik, K.A. (2004). Phosphorus uptake and growth promotion of chickpea by co-inoculation of mineral phosphate solubilizing bacteria and a mixed rhizobial culture. Aust J. Exp Agric. 44, 623-628.
- 19. H.G. Schlegel and B. Bowien (Eds.). Springer, Germany, 283-287.
- 20. Harrison, A.P. (1984). The acidophilic Thiobacilli and other acidophilic bacteria that share their habitat. Annu. Rev. Microbiol., 38, 265-292.
- 21. Haselwandter, K., and Bowen, G.D. (1996). Mycorrhizal relations in trees for agroforestry and land rehabilitation. Forest Ecology and Management, 81, 1-17.
- 22. Hilda, R., and Fraga, R. (2000). Phosphate solubilizing bacteria and their role in plant growth promotion. Biotech Adv., 17, 319-359.
- 23. Huber, R. and Stetter K. O. (1999). Aquificales. In: Embryonic ELS, No. 785998. Macmillan, Houndmills, England.1-7.
- 24. Igual, J.M., Valverde, A., Cervantes, E., and Velázquez E. (2001). Phosphate-solubilizing bacteria asinoculants for agriculture: use of updated molecular techniques in their study. Agronomie. 21, 561-568.
- 25. Illmer, P.A., and Schinner, F. (1995). Solubilization of inorganic calcium phosphates solubilization mechanisms. Soil BiolBiochem., 27, 257-263.
- 26. Imhoff, J.F., Siiling J., and Petri R. (1998). Phylogenetic relationship and taxonomic reclassificiation of *Chromatium* species and related purple sulfur bacteria. Int. J. Syst. Bacteriol., 48, 1029-1043.
- 27. Kapoor, K.K., and Mishra M.M., (1989).Microbial transformation of sulphur and plant nutrition. In: Soil microorganisms and crop growth. L.L. Somani and S.L. Bhandari. (Eds.). DiyajyotiPrakasam, India, 1-30.
- 28. Kelly, D.P., and Smith N.A., (1990). Organic sulfur compounds in the environment. Adv. Microb. Ecol., 11, 345-385.
- 29. Kelly, D.P., and WoodA.P., (2000). Reclassification of some species of *Thiobacillus*to the newly designated genera *Acidithiobacillus*gen. nov., *Halobacillus*gen. nov.and*Themithiobacillus*gen. nov. Int. J. Syst. Evol. Microbial., 50, 511-516.
- 30. Kelly, D.P., McDonald I.R., and Wood A.O. (2000). Proposal for the reclassification of *Thiobacillusnovellus* as *Starkeya novella* gen. nov., comb. nov., in the -subclass of the Proteobacteria. Int. J. Syst. Evol. Microbiol., 50, 1797-1802.
- 31. Khan, M.R., and Khan S.M. (2002). Effect of root-dip treatment with certain phosphate solubilizing microorganisms. Bioresource Technology, 85 (2), 213-215.
- 32. Khan, M.S., Ahmed, M., Oves, M., and Wani, P.A. (2010). Plant growth promotion by phosphate solubilizing fungi-current perspective. Arch. Agron. Soil Sci., 56, 73-98.
- 33. Khatbi, R. (2011). Using sulfur oxidizing bacteria and P solubilizing for enhancing phosphorous availability to Raphanussativus. African journal of plant science, 5(8), 430-435.

- 34. Khiari, L., and Parent, L.E. (2005). Phosphorus transformations in acid light-textured soils treated with dry swine manure. Can J. Soil Sci., 85, 75-87.
- 35. Kondratieva, E.N.(1989). Chemolithotrophy of phototrophic bacteria. In: Autotrophic bacteria.
- 36. Kuenen, J.G., and Beudeker, R.F. (1982).Microbiology of *Thiobacilli* other sulfur oxidizing autotrophs mixotrophs and heterotrophs.In: sulfur bacteria. J.P. post gate and D.P. Kelly (Eds.) University press, Cambridge, 473-497.
- 37. Mishra, A. (1991). Strategies for reclamation of mine areas through help of selected microbes, Symp.on strategies for ecosystem conservation, Bot. Section, 78 ISCA session, 64 (Abst.).
- 38. Nahas, E. (1996). Factors determining rock phosphate solubilization by microorganism isolated from soil. World J. Microb Biotech., 12, 18-23.
- 39. Rajan, S. (2002). Comparison of phosphate fertilizers for pasture and their effect on soil solution phosphate. Comm. Soil Sci. Plant Anal., 33, 2227-2245.
- 40. Reddell, P., Gordon, V., Hopkins, M.S. (1999). Ectomycorrhizas in *E. tetrodonta* and *E. miniata* in forest communities in tropical and their Role in rehabilitation of these forest following mining. Australian J. of Botany, 47, 881-907.
- 41. Rengel, Z., and Marschner, P. (2005). Nutrient availability and management in the rhizosphere: exploiting genotypic differences. New Phytology, 168, 305-312.
- 42. Rokhzadi, A., Asgharzadeh, A., Darvish, F., Nourmohammadi, G., and Majidi E. (2008). Influence of plant growth-promoting rhizobacteria on dry matter accumulation and yield of chickpea (*Cicerarietinum*L.) under field condition. Am-Euras. J. Agric. Environ. Sci., 3(2), 253-257.
- 43. Saber, K., Nahla, L.D., and Chedly, A. (2005). Effect of P on nodule formation and N fixation in bean. Agron. Sustain Dev, 25, 389-393.
- 44. Setiadi, Y. (2002). Mycorrhizal inoculum production technique for land rehabilitation. Journal of Tropical Forest Management, 1, 51-64.
- 45. Sharma, M.P., Bhatia, N.P., Chauhan, R.K.S., Adholeya, A. (2001). A Mycorrhizal dependency and growth responses of *Acacia nilotica* and *Albizzialebbeck*to inoculation by indigenous AM fungi as influenced by available soil P levels in a semi-arid Alfisol wasteland. New Forests, 21 (1), 89–104.
- 46. Sharma, M.P., Bhatia, N.P., Gaur, A., Adholeya, A. (1997). Mycorrhizal dependency of Acacia nilotica and Eucalyptus tereticoris to inoculation of indigenous VA Mycorrhizal fugal consortium in marginal wasteland soil. In proceedings XIth World Forestry Congress, 13-22 October 1997. Antalya, Turkey.
- 47. Sheoran, V., Sheoran, A, S., and Poonia, P. (2009). Phytominining: A review.Minerals Engineering, 22 (12), 1007-1019.
- 48. Siefert, E., and Pfennig N. (1979). Chemoautotrophic growth of *Rhodopseadomonus* species with hydrogen and chemotrophic utilization of methanol and formate. Arch. Microbiol., 122, 177-182.

- 49. Sorokin, D. Y., Lysenko A. M., Mityushina L., and Kuenen J. (2001). *Thioalkalimicrobiumaerophulum*gen. nov., sp. nov.and *Thioalkalimicrobiumsibericum*sp. nov. and *Thioalkalivibrioversutus*gen. nov., sp. nov.and *Thioalkalivibrionitratis*sp. nov.and *Thioalkalivibriodenitrificans*sp. nov., novel obligatelyalkalophilic and obligatelychemolithoautotrophic sulfur oxidizing bacteria from soda lakes. Int. J. Syst. Evol. Microbiol., 51, 565-580.
- 50. Stamford, N.P., Silva, J.A., Freitas, A., Araujo, Filho J.T. (2002). Effect of sulfur inoculated with Acidithiobacillus in a saline soil grown with Leucena and mimosa tree legumes. Biores. Technol., 81, 53-59.
- 51. Strohl, W.R. (1989). Genus I. *Beggiatoa*. In: Bergey's manual of systematic bacteriology, vol 3. J. T. Staley, M.P. Bryant, N. Pfennig and J.G. Holt (Eds.). Williams and Wilkinson, Baltimore, Md., 2091-2097.
- 52. Subbarao, N. S. (1988). Phosphate solubilizing microorganism. In: Biofertilizer in agriculture and forestry. Regional Biofert.Dev. Centre, Hissar, India.133-142.
- 53. Vernadskii, V.I., (1927). Istoriyamineralovzemnoikory (History of the minerals of the earth crust), 1, (2) (see Izbrannye Trudy, Vol. 3, Izd. An SSSR. 1955).
- 54. Vessey, J.K. (2003). Plant growth promoting rhizobacteria as biofertilizers. Plant Soil, 255, 571-586.
- 55. Vidyalakshmi, R., and Paranthaman, R. et al. (2009). Sulfur oxidizing bacteria and pulse nutrition- a review. World journal of agricultural sciences, 5 (3), 270-278.
- 56. Waksman, S.A. (1932). Principles of soil microbiology.2nd edition. Williams and Williams Co., Baltimore.
- 57. Whitelaw, M.A. (2000). Growth promotion of plants inoculated with phosphate solubilizing fungi. Adv Agron., 69, 99-151.
- 58. Zhou, K., Binkley, D., and Doxtader, K.G. (1992). A new method for estimating gross phosphorous mineralization and immobilization rates in soils. Plant soil, 147, 243-250.